Data preprocessing and integration for reproducible multiomic biomarker discovery and validation

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- Carefully designed study (begin with end in mind):
 - Targeted data collection
 - Include 'orthogonal' data types

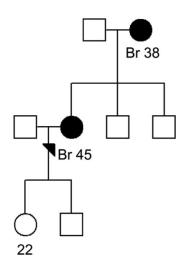
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 - Data preprocessing and integration
 - Discovery, development, optimization (and adaptation!)

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 - Mechanistic or functional validation
 - Additional genomic profiling?

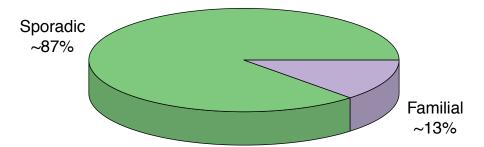
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- 4 Last but not least: Transparency!



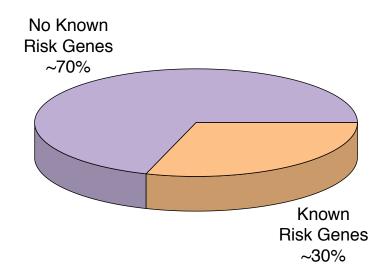
Breast Cancer Often Runs in Families



Breast Cancer

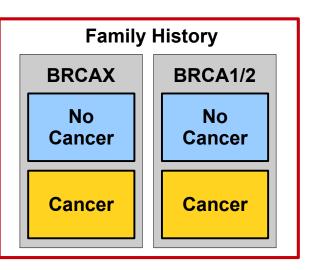


Familial Breast Cancer



Risk Subpopulations

No Cancer Cancer



Predicting Hereditary Breast Cancer

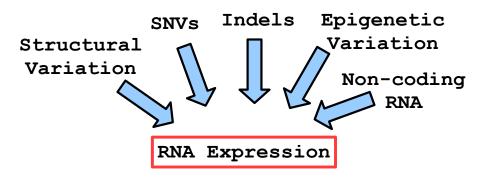
Goal: Develop a non-invasive biomarker for hereditary breast cancer risk

Key hypotheses or information to build our biomarker:

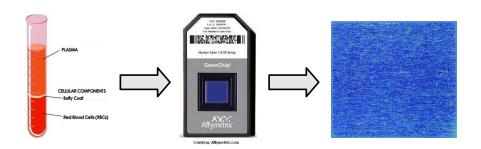
- Rare or highly penetrant genetic variation impacting key pathways (e.g. DNA repair)
- Germline-driven mRNA degregulation as an intermediate risk phenotype for familial breast-cancer susceptibility
- Expression patterns in peripheral blood will enable prospective identification of high-risk women who will develop breast cancer



RNA Expression as a Variation Surrogate



Microarray Profiles of Peripheral Blood



Patient Cohorts



Location	Туре	Patients
Utah	Retrospective	124
Ontario	Retrospective	36
Ontario	Prospective	37



Study Design Summary

Important elements of the study design:

- Study patients recruited and data collected specifically for this study
- Validation on a heterogenous population/dataset
- Clear purpose for the study and 'success' is predefined
- Collected expression array and DNA sequencing data (validation? multi-omic biomarker?)

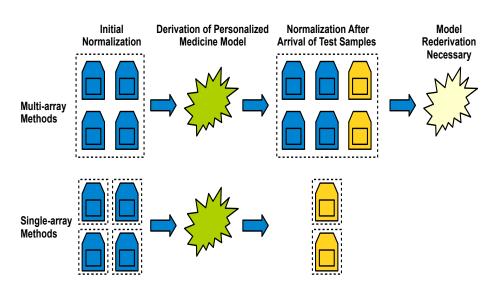
Goals of Personalized Genomic Medicine



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Data Normalization Methods

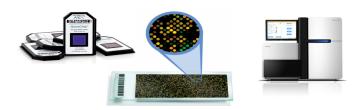


Microarray/Sequencing Normalization Methods

- Multisample Methods:
 - dChip (Li, PNAS, 2001)
 - QQ/RMA (Bolstad, Bioinformatics, 2003; Irizarry, Biostatistics, 2003)
 - RNA-seq: Conditional-QQ (Hansen, Biostatistics, 2012)
- Single Sample Methods:
 - MAS5 (Hubbell, Bioinformatics, 2002)
 - fRMA (McCall, Biostatistics, 2010)
 - Barcoding (Irizarry Nat Meth 2009; McCall, NAR, 2012)
 - RPKM/FPKM (Mortazavi, Nat Meth, 2008; Trapnell, Nat Biotech 2010)
- Single Array and Sequencing Integrative Methods:
 - SCAN-UPC (Piccolo, Genomics, 2012)



SCAN-UPC: Single Sample of Expression Estimates

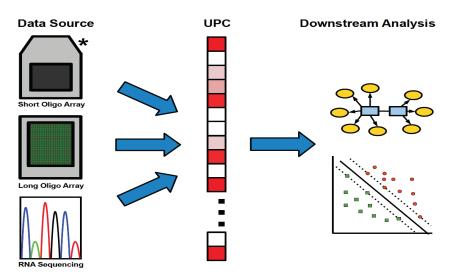


Our approaches: Single Channel Array Normalization (SCAN), and Universal Probability of Expression Codes (UPC)

- Uses background within a single sample
- Individual patient samples without any extraneous data
- Can be applied to all platforms: one and two color arrays, RNA-seq (UPC)
- Naturally combines data across platforms (UPC)

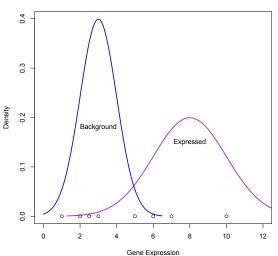


Biomarker and Personalized Medicine Workflows

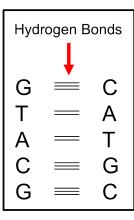


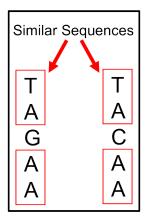
SCAN-UPC: Single Sample of Expression Estimates

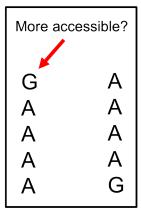
Two Component Gassian Mixture Model



SCAN-UPC Model Justification:







For Affymetrix arrays:

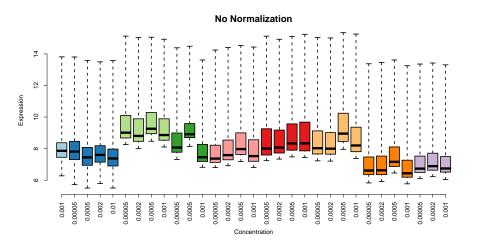
• Each component is $N(X\theta_m, \sigma_m^2)$ (m = 1, 2), where

$$x_i \theta_m = \alpha_m n_{iT} + \sum_{j=1}^{25} \sum_{k \in \{A,C,G\}} \beta_{jkm} I_{ijk} + \sum_{l \in \{A,C,G,T\}} \gamma_{lm} n_{ik}^2,$$

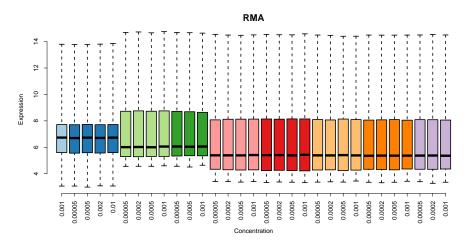
(Johnson et al., PNAS, 2006)



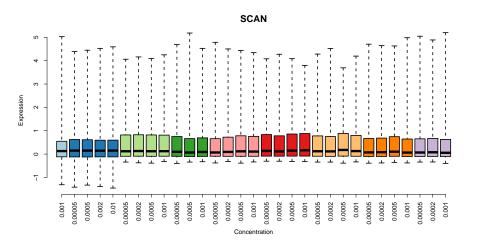
Batch and Design Effects



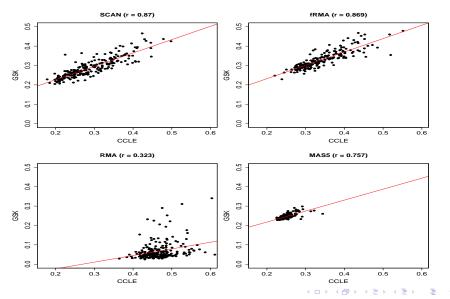
RMA



SCAN → Consistent Across Array Designs



Single Channel Array Normalization (SCAN)



For two-color arrays:

Suppose

$$\log(\mathbf{Y_i}) = (\log(Y_{i1}), \log(Y_{i2})) \sim N(\mathbf{m_k}, \Sigma_k)$$

where k is the G+C of the probe

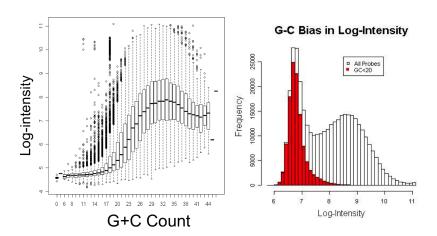
Transform to mean center and remove chip and dye effects:

$$\mathbf{Z_i} = \hat{\Sigma}_k^{-1/2} (\log(\mathbf{Y_i}) - \hat{\mathbf{m}}_k)$$

(Song et al., Genome Biology, 2007)

Apply a simple two-component mixture model

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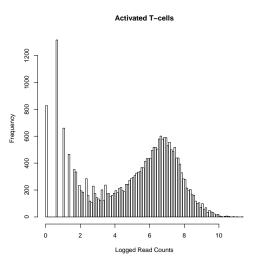


RNA-seq data:

- Mapping errors, repetitive regions
- 'Leaky' transcription
- Each component is $N(X\theta_m, \sigma_m^2)$, where

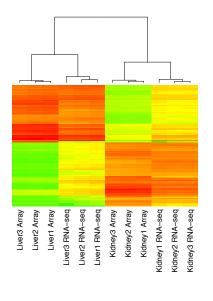
$$x_i\theta_m = \alpha_m + GC_i\beta + Len_i\gamma$$

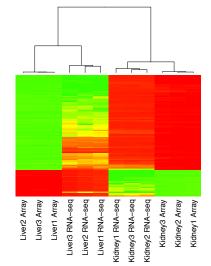
RNA-Seq Data:



(a) Normalized Array, Read Count

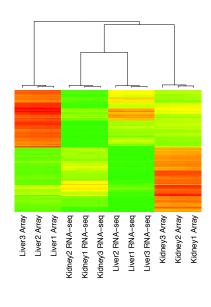
(c) UPC Array and Seq

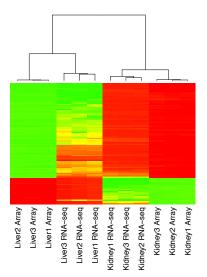




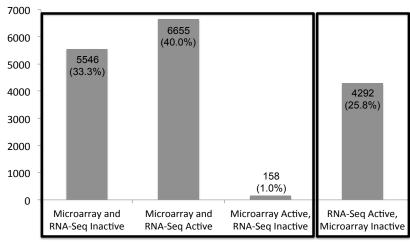
(b) Normalized Array, RPKM

(c) UPC Array and Seq



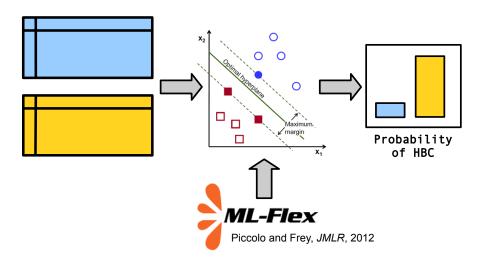


Active and Inactive Genes Across Platforms

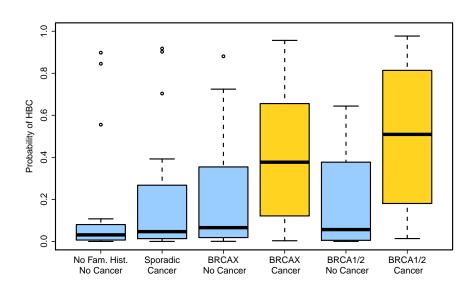


Microarray active or RNA-Seq inactive subset: 98.7% correspondence across platforms Discordant due to increased sensitivity of RNA-Seq

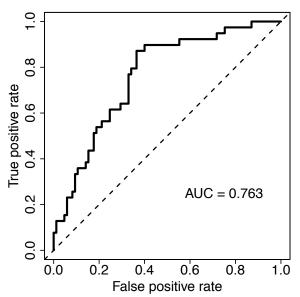
Risk Prediction



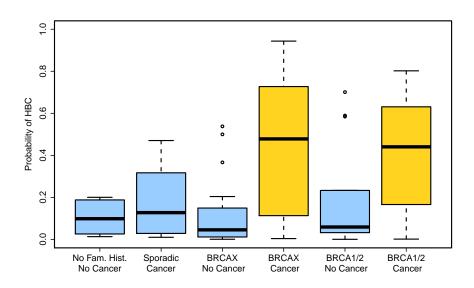
Utah – Predicted Risk



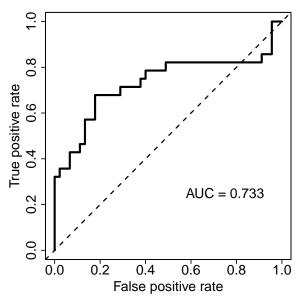
Utah — ROC Curve



Ontario – Predicted Risk



Ontario — ROC Curve

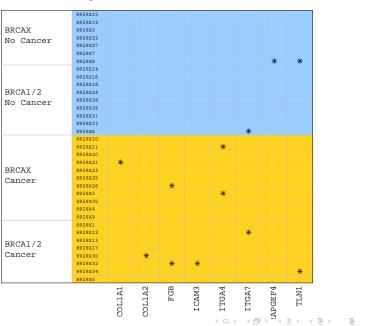


Top Pathway Results

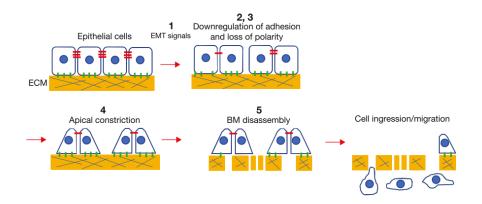
Pathway	AUC	Controls Mutated	Cancer Mutated
Integrin cell surface interactions	0.687	3/16	9/19
Cell adhesion molecules	0.682	2/16	8/19
PI3K Signaling System	0.676	4/16	10/19
Citrate/Krebs cycle	0.678	0/16	7/19
Fructose and mannose metabolism	0.668	1/16	7/19
ERBB signaling pathway	0.658	3/16	7/19

Pathways that performed well in both analyses are known to play a role in tumor development!

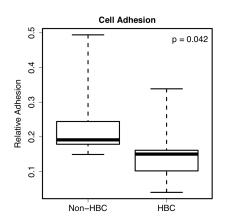
Integrin cell-surface interactions

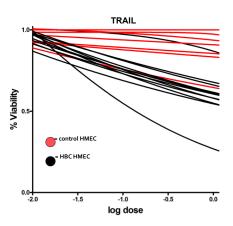


Epithelial Tissue Adhesion



Functional Results





Acknowledgements

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